33)-1 Parkinson's disease

Eur Respir J 2000; 16: 947-950 Printed in UK - all rights reserved SB 203580 inhibits therelease of TNF-d

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Effect of the p38 kinase inhibitor, SB 203580, on sephadex induced airway inflammation in the rat

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Effect of the p38 kinase inhibitor, SB 203580, on sephadex induced airway inflanmation in the rat. M. Birrell, D. Hele, K. McCluskie, S. Webber, M. Foster, M.G. Belvist. ©ERS Journals Ltd 2000.

ABSTRACT: SB 203580 is a pyridinyl imidazole compound which inhibits the release of pro-inflammatory cytokines, such as tumour necrosis factor-cd. (Th/Fe;q) and intereleukin-1B (IL-1B), in vitro and in vivo by inhibiting p38 mitogen-activated protein kinase (MAFK).

The present study investigated the effects of SB 203580 in a model of airway immuniton induced by the topical administration of Sephades into the rat airways. This inflammatory response is characterized by the development of lung oedema, airway tissue inflammatory cell recruitment and an increase in lung TNF- α and IL-1 β levels.

Les Sephades-induced lung oedema was accompanied by a significant increase in lung tissue TNF-0 but not IL-19 levels. There was also a significant increase in lung tissue macrophages and an increase in essinophils which did not reach significance. SB 203589 administration significantly inhibited lung oedema (EDS0-18 mg/kg³) in a dose-related manner but was without significant effect on lung tissue cell recruitment or cytokine levels.

These data suggest that the increase in tumour necrosis factor-\alpha and lung oedema are separate processes which both contribute to Sephadex pathology. Furthermore, the inhibitory effect of SB 203580 on Sephadex-induced lung oedema suggests that p38 kinase inhibitors may be of use in pulmonary pathologies in which lung oedema is a feature.

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The Sephadex model of lung oedema in the rat is a model characterized by acute alveolitis and bronchiolitis, leading to inflammatory cell infiltration and interstitial oedema. In fact this model appears to parallel many of the pathophysiological features associated with human interstitial lung diseases [1]. This model has been used previously to demonstrate the anti-inflammatory properties of compounds such as glucocorticosteroids and ebselen hased upon their ability to produlate lung oedems [2, 31].

based upon their ability to modulate lung oedema [2, 3]. SB 203580 (44.4 fluorophenyl).2-(4-methylsulphinyl-phenyl)-5-(4-pyridyl) imidazole) is a pyridinyl imidazole compound which inhibits interleukin (IL)-1β and tumour necrosis factor (TNF-α) production from human monocytes [4]. The molecular target of this and related compounds has been identified as p38 mitogen-activated protein kinase (MAPK) [4]. In addition to in vitro data, p38 kinase inhibitors, such as SB 203580, have been shown to inhibit LPS-induced systemic TNF-α release in rats [5, 6] and has been shown to be efficacious in in vivo models of arthritis [5] and inflammatory angiogenesis [7]. Furthermore, it has recently been demonstrated that in the airways SB 203580 inhibits bronchoalveolar (BAL) IL-1β levels and the associated neutrophilia in an LPS-induced model of airway inflammation in the rat [6] and BAL TNF-α levels following allergen challenge in sensitized Brown Norway rats [8].

Increased levels of TNF-or in the airways following Sephadex treatment have been suggested to play a causative role in Sephadex-induced lung pathology [9]. In this study the effect of SB 203580 on Sephadex-induced airway inflammation in the rat was examined.

Materials and methods

Animals

Male, Sprague-Dawley rats (330–350 g) were purchased from Harian-Olac (Bicester, UK) and housed for 1 week before initiating experiments. Food and water were supplied ad libitum. UK Home Office guidelines for animal welfare based on the Animals (Scientific Procedures) Act 1986 were strictly observed.

Methods

Rats were dosed intratracheally (i.i.) with vehicle (saline) or Sephadex beads (5 $\operatorname{mg/kg}^{-1}$) at a dose volume of 1 $\operatorname{ml.kg}^{-1}$ under halothane anaesthesia (4% in oxygen for 3 min). SB 203580 (1, 3, 10, 30 & 100 $\operatorname{mg/kg}^{-1}$) or vehicle (1% catboxymethyl cellulose (CMC) in distilled water, 2

mL·kg⁻¹) were administered orally 1 h prior to and 5 h post Sephadex administration. The positive standard, dexamethasone (1 mg·kg⁻¹) was dosed orally 24 and 1 h prior to Sephadex.

Measurement of lung oedema

Rats were killed 24 h post-Sephadex by sodium pentobarbitone (200 mg kg⁻¹; h₋) overdose, the heart and lungs were removed en bloc, and the lung wet weights determined and expressed per 100 g initial body weight Percentage inhibition of oedema compared to the Sephadex i.I/wheiled p-0, control group was then determined for each treatment group. A dose-response curve was generated, a sigmoidal fit obtained for the data and an EDSo value (dose that produced 50% of the maximum inhibition of lung oedema) calculated.

Measurement of cytokine protein levels in the lung

After weighing the rat lung, approximately 500 mg of the lung tissue was chopped and flash frozen in liquid nitrogen and stored at -20°C until needed. Approximately 250 mg of lung tissue was weighed and homogenized with 1 mL of ice cold saline. The homogenized sample was then spun at 800 g for 10 min. The resulting supernatant was taken off and stored at -20°C. TNF-α and IL-1β levels in the lung tissue supernatant were determined using a rat specific enzyme-linked immunosorbent assay (ELISA).

Measurement of cell recruitment in the lung tissue

After weighing the rat lungs for oedema determination. one lobe of the lung was removed and perfused with ice cold Roswell Park Memorial Institute medium (RPMI) to remove any blood and then finely chopped. The amount and type of white blood cells in the lung tissue were determined by incubating the chopped lung tissue with 10 mL of RPMI containing collagenase (1 mg·mL⁻¹) and DNase (25 µg·mL⁻¹) in a shaking water bath at 37°C for 1 h. The recovered cells were then passed through a 70 µm cell sieve and spun at 800 g for 10 min at 4°C. The supernatant was discarded, 10 mL of RPMI added and the pellet resuspended. The pellet washing step was carried out twice to remove the collagenase and DNase. After the final wash the pellet was resuspended in 1 mL of RPMI. Total white blood cell counts were determined on an automated cell counter (Cobas Argos, Roche ABX Hematologie, Montpellier, France). Cytospins of these samples were prepared by centrifugation of 100 uL aliquots in a cytospin (Shandon, Runcorn, UK) at 800 g for 5 min at room temperature. Slides were fixed and stained on a Hema-tek 2000 (Ames Co., Elkhart, USA) with modified Wrights-Giernsa stain. Four part differential counts on 200 cells per slide were performed following standard morphological criteria and the percentage of eosinophils, macrophages/monocytes, lymphocytes and neutrophils were determined [10].

Materials

SB 230580 (4(4-fluorophenyl)-2(4-methylsulphinylphenyl)-5(4-pyridyl) imidazole) was synthesized by the Medicinal Chemistry Department at Aventis Pharma (Dagenham, Essex, UK). Euthatal and halothane were obtained from Rhône Merieux (Harlow, UK) and RPMI from GIBCO (Paisley, UK). Roche Diagnostics (Lewes, UK) supplied the Dhase and collagenase. Sephadex G-200 was purchased from Pharmacia (Uppsala, Sweden). CMC and dexamethasone were purchased from Sigma (Poole, UK). TNF-α ELISA kits were from R&D systems (Abingdon, UK) and IL-1β ELISA kits were from Lifescreen (Watford, UK).

Analysis

All values are presented as mean±stM from n=8 rats per group. The percentage inhibition of oedema (compared to the Sephadex administered, vehicle-treated group) was determined for each treated group. The dose-response curve for inhibition of lung oedema by SB 203580 was calculated by least squares, non-linear iterative regression with the "PRISM" curve fitting programme (Graphpad Instat software programme, San Diego, CA, USA). An EDSO value (dose of SB 203580 required to produce 50% of the maximum inhibition) was subsequently interpolated from a curve of best fit. The data were analysed using the Kruskal-Wallis nonparametric test with a correction for multiple comparison's using Dunnert's critical values. A p-value of less than 0.05 was considered to be statistically significant (**: p>0.05 and ***: p>0.011.

Results

Lung oedema

Sephadex instillation evoked a significant oedema (21%, p<0.01). SB 203580 produced a dose-dependent inhibition of lung oedema (ED50 of 18 mg/kg²) when compared to Sephadex-instilled, vehicle-treated animals (fig. 1). Dexamethasone (1 mg/kg²) significantly reduced lung wet weight (117% inhibition, p<0.01).

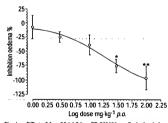


Fig. 1. – Effect of the p38 inhibitor, SB 203580, on Sephadex-induced lung codema Percentage inhibition of Sephadex-induced lung codema by SB 203580 (1–100 mg/sg², administered 1 h before and 5 h after Sephadex instillation, ρ.ο.). Points represent meansem of 8 rats. ED30=18 mg/sg². *: p=c0.0; **: p=c0.0;

Cytokine protein levels in the lung tissue

Sephadex treatment significantly increased lung tissue TNF- α levels 150.6427.2 to 1104.7 ± 243 9 pgg lung tissue.1 (733%, p<0.01) but had no impact, in this study, on IL-1 β levels in the lung tissue. SB 203580, at the doses tested, did not significantly impact on either cytokine, whereas dexamethasone significantly reduced both TNF- α (from 1104.7 ± 24.3) to 207.95 ± 20.91 pgg lung tissue.1 pc.0.01) (figs. 2a and b).

Inflammatory cell content in the lung tissue

Sephadex instillation caused an increase in lung tissue macrophages (5.2±0.6 to 10.5±1.9 cell 10³ grissue¹, p=0.05) and ossinophils (0.79±0.26 to 2.39±0.78 cell × 10³ grissue¹, p=0.05), however, neutrophils were unaffected. SE 203580 reatment did not significantly affect inflammatory cell number, whereas dexamethasone significantly reduced eosinophil number (2.39±0.78 to 0.22±0.035 cell × 10³ grissue¹, p=0.01), neutrophil number (7.71±1.95 to 2.75±0.30 cell × 10³ grissue¹, p=0.01) and macrophage number (10.5±1.9 to 1.8±0.2 cell × 10³ grissue³, p=0.01).

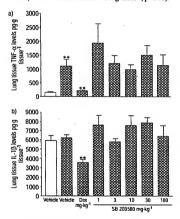


Fig. 2. – Effect of the p.18 inhibitor, SB 201580, on Sephadex-induced volutile production in lung tissue. Effect of SB 20580 (1-100 ng/s), ρ.ο.) on Sephadex-induced tumour necrosis factors—α (a) and interleukin-19 (b) levels and in lung tissue as measured by rat specific enzyme-linked immunosorbent assay. Data are expressed as the meansatus of 8 rats, Dec. deamerbationer, THF-Cc tumour necrosis the meansatus of 8 rats, Dec. deamerbationer, THF-Cc tumour necrosis challenge. Statistical significance was assessed by Knickal Walls with a correction for Dumertic strictical values, (**p. Do.05, ***p. p.0.0105, ***p. p.0.0105, ***p.0.0105, **p.0.0105, ***p.0.0105, **p.0.0105, ***p.0.0105, ***p.0.0105, ***p.0.0105, ***p.0.0105, **

Discussion

Recently, attention has focused on the MAPK superfamily due to their consistent activation by pro-inflammatory cytokines and to their role in nuclear signalling. Interest in one of these families, the p38 family, has been particularly intense following the discovery that p38 inhibitors are anti-inflammatory in in vivo models of arthritis and inflammatory angiogenesis [4, 5, 7]. However, the role and the contribution of p38 kinase in airway inflammation is less clear. In this study the effect of SB 203580 on Sephadex-induced airway inflammation has been investioared.

SB 203580 significantly inhibited Sephadex induced lung oedema in a dose-dependent manner achieving an ED50 of 18 mg.kg⁻¹. The potency of SB 203580 in this model agrees with data obtained in models of arthritis in mice and rats where the effect is attributed to inhibition of p38 kinase [5]. Although a nonspecific action of the compound, unrelated to its inhibitory action on p38 kinase, cannot be ruled out. Alternatively, SB 203580 failed to impact on inflammatory cell recruitment into lung tissue or on TNF-α and IL-1β levels in lung tissue. However, the steroid, dexamethasone, which was used as a positive control completely abrogated Sephadexinduced lung oedema, cell recruitment into the lung tissue and TNF-\alpha levels in the lung tissue. The inability of SB 203580 to inhibit the Sephadex-induced increase in TNF-α levels in the lung tissue cannot be attributed to the bioavailability and tissue distribution of the compound since the compound did have an inhibitory action on lung oedema. Previously the authors have shown that SB 203580 inhibits LPS-induced systemic TNF-α release but not aerosolized LPS-induced airway TNF-α release into BAL fluid and the associated initial phase of neutrophilia obtained in this model [6]. Therefore, the inability of SB 203580 to impact on airway tissue TNF-α levels after Sephadex is not such a surprising finding. However, this phenomenon appears to be stimulus specific since SB 203580 has been shown to inhibit the increase in BAL TNF-α levels following allergen challenge in sensitized

Brown Notway rats [8]. The reasons for the lack of effect of SB 203580 on TNF-α levels are not clear but could be related to the differential expression and sensitivity of the different p38 isoforms to inhibition by SB 203580, SB 203580 has been shown to inhibit p38-α and p38-β but not p38-γ and p38-δ [11]. Thus, if p38-γ and p38-δ are the predominant isoforms involved in the release of TNF-α in the lung tissue following Sephadex this could explain the apparent ineffectiveness of SB 203580. Alternatively, it may be that the release of TNF-α may not be solely controlled via the p38 kinase signal transduction pathway and could be to activation of other signalling pathways. In fact, both p38 and ERK are involved in LPS-induced TNF-α production from macrophages in vitro [12].

Sephadex-induced inflarimatory cell recruitment into the rat lung was not inhibited by SB 203580. In contrast to these observations, the authors have recently reported that SB 203580 inhibited an LPS-induced increase in neutrophils and IL-JB in the lung tissue [6]. However, the inflammatory cell and cytokine release profile in the Sephadex model is completely different to that induced by aerosolized LPS. In this study, however, the stimulus

used to produce the inflammatory response was Sephadex rather than LFS, and Sephadex did not evoke an increase in neutrophils and IL-1β, therefore, the effect of SB 203580 on this axis could not be tested. In fact, this observation is consistent with a previous study by the authors where TNF-α, endothelin-1 but not IL-1β levels were elevated in BAL fluid from rats 24 h following i.t. Sephadex treatment [3].

To conclude, these data suggest that the increase in lung tissue numour necrosis factor-Q and lung oddema are separate processes which may both contribute to Sephadev pathology and that the development of lung codema after Sephadev is not dependent on the release of this system. However, it is possible that inhibitors of 938 kinase may be responsible for the reduction of Sephadev-induced lung codema and that inhibitors of this class may be of use in pulmonary pathologies in which lung codema is a

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